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Linker-Based Hemisuccinate Derivatives of Artemisinin: Synthesis and Antimalarial Assessment against Multidrug-Resistant *Plasmodium yoelii nigeriensis* in Mice¹

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(5) Supporting Information

ABSTRACT: Artesunic acid 5, the hemisuccinate derivative of dihydroartemisinin 2, is the only clinically useful water-soluble derivative of artemisinin 1. However, being a lactol ester, it is rapidly hydrolyzed back to dihydroartemisinin in aqueous alkaline solution, a reaction that seriously limits its utility. A new series of potentially more stable linker-based hemisuccinate derivatives 12a-i and 14a-c have been prepared. The process involved acid-catalyzed reaction of dihydroartemisinin with various diols and polyethylene glycols to give hydroxy-functionalized ethers 7a-i and 10a-c and their further derivatization to hemisuccinate esters 12a-i and 14a-c. Both the hydroxy-functionalized ethers 7a-i and 10a-c and their further derivatization to hemisuccinate esters 12a-i and 14a-c. Both the hydroxy-functionalized ethers 7a-i and 10a-c and their hemisuccinate derivatives 12a-i and 14a-c have been assessed for antimalarial activity against multidrug-resistant *Plasmodium yoelii nigeriensis* in Swiss mice. Several of these



hemisuccinate derivatives have shown very promising activity. Hemisuccinate derivatives 12f and 12i, the two most active compounds of the series, provided 100% protection to malaria-infected mice at 24 mg/kg \times 4 days and therefore are twice as potent as artesunic acid, which provides a similar level of protection at 48 mg/kg \times 4 days.

INTRODUCTION

Malaria is a major parasitic disease of the tropical and subtropical countries including India.² Nearly 300-500 million episodes of malaria infections occur annually, killing more than a million people with severe and cerebral malaria. The malaria problem has been further complicated with the emergence of malarial parasites resistant to the commonly used antimalarial drugs.³ Chloroquine-resistant Plasmodium falciparum is present in most of the countries of Asia, Africa, and South America. Resistance to the sulfonamide-pyrimethamine combination is widespread in southeast Asia and South America. Field trials with mefloquine have met with rapid emergence of malarial parasites resistant to the drug. Resistance to quinine is not common, but the duration of the treatment with the drug is long and requires hospitalization. Currently, artemisinin 1 and its derivatives dihydroartemisinin 2, artemether 3, arteether 4, and artesunic acid 5 (Figure 1) are the only class of drugs that are effective against multidrug resistant malaria.⁴ Artesunic acid, the hemisuccinate ester of dihydroartemisinin 2, is the only clinically useful water-soluble derivative of artemisinin. However, being a lactol ester, it is rapidly hydrolyzed back to dihydroartemisinin in aqueous alkaline solution. Furthermore, dihydroartemisinin, being highly sensitive to basic conditions at ambient temperature, degrades into nonperoxide-containing compounds. This rapid hydrolysis of artesunic acid in aqueous solution greatly restricts its use, which reflects the need for more stable and possibly more effective water-soluble substitutes of artesunic acid. To meet these objectives, we have prepared a series of hydroxyfunctionalized ether derivatives of dihydroartemisinin 7a–i and 10a–c and converted them into their hemisuccinate derivatives 12a–i and 14a–c, which readily form watersoluble sodium salts when treated with aqueous sodium bicarbonate.⁶ Hemisuccinate derivatives 12f and 12i, the two most active compounds of the series, provided 100% protection to malaria-infected mice at 24 mg/kg × 4 days. Artesunic acid provides a similar level of protection at 48 mg/ kg × 4 days. Compound 12h was also assessed for its stability in aqueous bicarbonate and carbonate solution. It did not show any detectable degradation in 48 h at ambient temperature, while artesunic acid was completely degraded under these conditions.

CHEMISTRY

Dihydroartemisinin 2 was prepared from artemisinin 1 using the known procedure.⁷ BF₃·OEt₂ catalyzed reaction of 2 with alkane diols **6a–i** in CH₂Cl₂ at room temperature furnished the corresponding hydroxy-functionalized ether derivatives **7a–i** (β -isomer) and **8a–i** (α -isomer) in 31–67% yields as diastereomeric mixtures (approximately in the ratio of 3:1), with β -isomers as the major products. A similar reaction of 2 with polyethylene glycols **9a–c** furnished **10a–c** and **11a–c** in 49–55% yields (Scheme 1, Table 1). In most of the cases only β -isomers of these ether derivatives could be obtained in pure form and were used for the preparation of

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Figure 1. Artemisinin 1 and its clinically useful derivatives 2-5.

Scheme 1^a



^aReagents and conditions: (a) BF₃. OEt₂, CH₂CI₂, room temp, 15 h.

Table 1. Hydroxy-Functionalized Ether Derivatives 7a-i, 8a-i, 10a-c, and 11a-c



hemisuccinates.⁸ The hemisuccinate derivatives 12a-i and 14a-c were prepared by the treatment of hydroxy-

functionalized ethers 7a-i and 10a-c with succinic anhydride using either pyridine or Et_3N as a base and 4-N, Scheme 2^{*a*}



^aReagents and conditions: (a) pyridine or Et₃N, DMAP, CH₂CI₂, 0 °C, 1 h; (b) CH₂N₂, ether, 0 °C.

Table 2. Hemisuccinate Derivatives 12a-i and 14a-c



^{*a*}12g is a mixture of diastereomers, as it was prepared from a mixture of 7g and 8g.

N-dimethylaminopyridine (DMAP) as a catalyst in CH_2Cl_2 at 0 °C to room temperature (Scheme 2, Table 2). The hemisuccinate derivatives **12a–i** and **14a–c** were further converted to the corressponding methyl esters **13a–i** and **15a–c** to determine spectroscopic and analytical data (¹H NMR, ¹³C NMR, mass, and microanalysis).

ANTIMALARIAL ACTIVITY⁹

Artesunic acid, when given intramuscularly at 48 mg/kg \times 4 days, provides 100% protection to mice infected with multidrug-resistant *P. yoelli nigeriensis*. At 24 mg/kg \times 4 days, it does not provide any protection. Since the objective of the study was to select compounds exhibiting activity profiles

Table 3. Blood Schizontocidal Activity of Hydroxy Functionalized Ethers 7a-i and 10a-c against Multidrug-Resistant Strain *P. yoelii nigeriensis* in Swiss Mice via im Route^{*a*,9}

compd	log P	dose (mg/kg × 4 days)	% suppression of parasitemia on day 4 ^{b,c}	cured/treated	compd	log P	dose (mg/kg × 4 days)	% suppression of parasitemia on day 4 ^{b,c}	cured/treated
7a	2.99	48	100	12/12			3	100	0/6
		24	100	12/12	$7\mathbf{g} + \mathbf{8g}^d$	5.64	48	100	6/6
		12	100	9/11			24	100	5/5
		6	100	3/6			12	100	5/5
7b	3.10	48	100	5/5			6	100	1/5
		24	100	5/5			3	99.3	0/5
		12	100	9/11	7h	6.05	48	100	6/6
		6	100	6/12			24	100	5/12
		3	98.3	0/6			12	100	0/6
7c	3.55	48	100	5/5			6	100	0/6
		24	100	5/5	7i	6.89	48	100	4/12
		12	100	17/17			24	100	0/6
		6	100	11/12	10a	2.83	48	100	6/6
		3	99.5	0/6			24	100	6/6
7d	3.97	48	100	5/5			12	100	7/12
		24	100	5/5			6	100	0/6
		12	100	5/5	10b	2.68	48	100	2/6
		6	100	1/5			24	100	0/6
7e	4.38	48	100	6/6			12	100	0/6
		24	100	6/6	10c	2.52	48	100	8/10
		12	100	6/6			24	100	0/5
		6	100	9/12			12	100	0/5
		3	100	0/6	4	3.84	6	100	6/6
7f	5.22	48	100	6/6			3	100	0/6
		24	100	6/6	5	3.04	48	100	6/6
		12	100	6/6			24	100	0/5
		6	100	5/6			12	96.36	0/5

^{*a*}The drug dilutions of hydroxy functionalized ethers 7**a**–**i** and 10**a**–**c** were prepared in ground oil and administered to a group of five or six mice at each dose from day 0 to day 3, once daily. ^{*b*}Percent suppression = $[(C - T)/C] \times 100$, where *C* is the parasitemia in control group and *T* is the parasitemia in treated group. ^{*c*}100% suppression of parasitemia means no parasites were detected in 50 oil immersion fields during microscopic observation. ¹⁰ ^{*d*}Compounds 7**g** and 8**g** could not be separated and were tested as mixtures.

comparable to or better than that of artesunic acid, all newly prepared hemisuccinate derivatives 12a-i and 14a-c were initially screened at 48 mg/kg × 4 days by the im route. Compounds found active at 48 mg/kg × 4 days were further screened at lower doses. The results are shown in Table 4. Similar dose-dependent activity data were generated for hydroxy-functionalized intermediate compounds 7a-i and 10a-c. The results are depicted in Table 3.

RESULTS AND DISCUSSION

In the present study, the length of the side chain of the hydroxy-functionalized ethers was systematically increased and its effect on the antimalarial activity was assessed. Simultaneously, each of the hydroxy-functionalized ethers was converted to its hemisuccinate derivative and assessed for its antimalarial activity. Thus, two sets of SAR data were generated (Table 3 and Table 4).

Both 7a and 7b showed modest activity compared with arteether; both these compounds showed 100% protection at 24 mg/kg \times 4 days, while at 12 mg/kg \times 4 days, both compounds showed only partial protection. Arteether in this assay showed 100% protection at 6 mg/kg \times 4 days. Thus, these two compounds were significantly less active than arteether. Their respective hemisuccinate derivatives **12a** and **12b** also did not provide any significant protection at 48 mg/kg \times

4 days. Compounds 7c-e showed 100% protection at 12 mg/kg × 4 days and partial protection at 6 mg/kg × 4 days. However, none of their hemisuccinate derivatives 12c-e provided significant protection at 48 mg/kg. The activity of hydroxy-functionalized ethers did not show any improvement with further increase in the chain length. Both 7f and 7g showed similar levels of protection as provided by 7c-e. However, their hemisuccunate derivatives showed remarkable improvements in activity. While 12f showed 100% protection at 24 mg/kg, 12g showed a similar level of protection at 48 mg/kg × 4 days.

Further increase in the chain length resulted in reduced activity of the hydroxy-functionalized ethers. Both 7h and 7i were considerably less active than 7c-f. Their hemisuccinate derivatives, however, did not show similar drops in activity. While **12i** provided 100% protection at 24 mg/kg, **12h** provided similar level of protection at 48 mg/kg \times 4 days.

Of the polyethylene glycol derivatives 10a-c, only 10a showed 100% protection at 24 mg/kg. Similarly, their hemisuccinate derivatives 14a-c showed poor activity; none of these compounds showed any significant protection at 48 mg/kg × 4 days.

Hemisuccinate derivative **12h** was also assessed for its stability in aqueous $NaHCO_3$ and aqueous Na_2CO_3 solution. It did not show any detectable degradation within 48 h at ambient

Table 4. Blood Schizontocidal Activity of Hemisuccinates 12a–i and 14a–c against Multidrug-Resistant Strain *P. yoelii nigeriensis* in Swiss Mice via im Route^{*a*,9}

compd	log P	dose mg/kg × 4 days	% suppression of parasitemia on day 4 ^{b,c}	cured/treated
12a	2.89	48	100	0/6
12b	2.99	48	100	0/6
12c	3.45	48	100	5/12
12d	3.86	48	94.91	0/6
		24	92.73	0/6
12e	4.28	48	98.55	0/6
12f	5.12	48	100	6/6
		24	100	6/6
		12	100	0/6
$12g^d$	5.53	48	100	6/6
		24	93.09	0/5
		12	67.27	0/5
12h	5.95	48	100	5/5
		24	100	8/11 + 1/5
		12	98.18	0/5
12i	6.78	48	100	5/5
		24	100	5/5
		12	71.27	0/5
		6	57.09	0/5
14a	2.73	48	100	1/11
14b	2.54	48	92	1/6
14c	2.42	48	97	0/6
5	3.04	48	100	6/6
		24	100	0/5
		12	96.36	0/5

^{*a*}The drug dilution of hemisuccinates **12a**–i and **14a**–c were prepared in 5% NaHCO₃ and administered to the group of five or six mice at each dose from day 0 to day 3, once daily. ^{*b*}Percent suppression = $[(C - T)/C] \times 100$, where C is the parasitemia in control group and T is the parasitemia in treated group. ^{*c*}100% suppression of parasitemia means no parasites were detected in 50 oil immersion fields during microscopic observation.¹⁰ ^{*d*}Compound **12g** is a mixture of diastereomers, as it is prepared from a mixture of 7g and 8g.

temperature, while artesunic acid was completely degraded under these conditions.

Thus, of the hemisuccinate derivatives, both 12f and 12i are superior to artesunic acid and both are twice as active as artesunic acid while 12g and 12h are comparable to artesunic acid. In addition, these compounds have the advantage of being more stable than artesunic acid in aqueous NaHCO₃ solutions.

CONCLUSION

We have prepared a new series of linker-based hemisuccinate derivatives of dihydroartemisinin, several of which have shown a high order of antimalarial activity against multidrug-resistant *P. yoelii nigeriensis* in Swiss mice. Hemisuccinate derivatives **12f** and **12i**, the most active compounds of the series, are twice as active as artesunic acid, the only clinically useful water-soluble derivative of artemisinin. Hemisuccinate **12h**, an active representative of the series, was also found to be stable in aqueous alkaline solution.

EXPERIMENTAL SECTION

General. All glass apparatuses were oven-dried prior to use. Melting points were taken in open capillaries on a Complab melting point apparatus and are presented uncorrected. Infrared spectra were recorded on a Perkin-Elmer FT-IR RXI spectrophotometer. ¹H NMR and ¹³C NMR spectra were recorded using a Bruker Supercon Magnet DPX-200 or DRX-300 spectrometer operating at 200 and 300 MHz, respectively, for ¹H and at 50 and 75 MHz, respectively, for ¹³C and at 400 MHz using CDCl₃ as solvent. Tetramethylsilane (δ 0 0.00 ppm) served as an internal standard in ¹H NMR, and CDCl₃ (δ 77.0 ppm) was the internal standard in ¹³C NMR. Chemical shifts are reported in parts per million. Splitting patterns are described as singlet (s), doublet (d), triplet (t), and multiplet (m). In NMR, numbering of atoms is presented according to the usual numbering in artemisinin as indicated in the text. Fast atom bombardment mass spectrometry (FABMS) data were obtained on JEOL SX-102/DA-6000 mass spectrometer using argon/xenon (6 kV, 10 mA) as the FAB gas. Glycerol or *m*-nitrobenzyl alcohol was used as matrix. Electrospray mass spectrometry (ES-MS) results were recorded on a MICROMASS QUATTRO II triple quadruple mass spectrometer. Elemental analyses were performed on a Vario EL-III C, H, N, S analyzer (Germany) and Carlo-Erba-1108 C, H, N elemental analyzer (Italian), and values were within $\pm 0.5\%$ of the calculated values; therefore, these compounds meet the criteria of ≥95% purity. Reactions were monitored on silica gel TLC plates (coated with TLC grade silica gel, obtained from Merck). Detecting agents used (for TLC) were iodine vapors, and/or spraying was done with an aqueous solution of vanillin in 10% sulfuric acid followed by heating at 150 °C. Column chromatography was performed over Merck silica gel (particle size, 60-120 mesh) procured from Qualigens (India) and flash silica gel (particle size, 230-400 mesh). All chemicals and reagents were obtained from Aldrich (U.S.), Lancaster (England), or Spectrochem (India) and were used without further purification. The log P values of the compounds were calculated using Chem Draw Ultra 10.0 software.

General Procedure for Preparation of Hydroxy-Functionalized Ethers 7a–i and 10a–c (Compound 7d as Representative). To a solution of dihydroartemisinin 2 (1.0 g, 3.52 mmol) and pentane-1,5-diol 6d (1.10 g, 10.5 mmol) in dry dichloromethane (20 mL) was added $BF_3 \cdot OEt_2$ (0.25 mL) at room temperature. The reaction mixture was stirred at the same temperature for 15 h. Then the reaction mixture was neutralized with saturated sodium bicarbonate solution (25 mL) and extracted with ether (3 × 25 mL). The combined organic layer was washed with water (2 × 25 mL) and then with brine (2 × 25 mL), dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The resultant crude product, upon column chromatography over silica gel using ethyl acetate/ hexane (15:85) as eluant, gave 7d (0.122 g) as a solid, a mixture of 7d and 8d (0.687 g), and 8d (0.037 g) as a solid, the combined yield being 65%.

7a. White solid; mp 98–100 °C; FT-IR (KBr, cm⁻¹) 3426.4, 2930.0, 2875.0, 1453.9, 1376.6, 1154.6, 1018.1, 757.5; ¹H NMR (400 MHz, CDCl₃) δ 0.93 (d, 3H, *J* = 7.3 Hz, CH₃), 0.97 (d, 3H, *J* = 6.7 Hz, CH₃), 1.23–2.05 (m, 10H), 1.44 (s, 3H, CH₃), 2.35–2.42 (m, 1H), 2.65–2.71 (m, 1H), 3.65 (m, 1H), 3.76 (m, 2H, <u>CH₂OH</u>), 3.91 (m, 1H), 4.86 (d, 1H, *J* = 3.4 Hz, C₁₀-H), 5.46 (s, 1H, C₁₂-H). FABMS (*m*/*z*): 335 [M + Li]⁺, 267 [M⁺ – OCH₂CH₂OH]. Anal. Calcd for C₁₇H₂₈O₆: C, 62.17; H, 8.59. Found: C, 62.20; H, 8.60.

Ethers 7**b**–**i** and 10**a**–**c** were prepared using the above procedure. 7**b**. White solid; mp 74–75 °C; FT-IR (KBr, cm⁻¹) 3461.4, 2927.3, 1594.5, 1351.4, 1102.2, 1017.0, 759.2; ¹H NMR (200 MHz, CDCl₃) δ 0.91 (d, 3H, *J* = 7.4 Hz, CH₃), 0.95 (d, 3H, *J* = 7.3 Hz, CH₃), 1.22–2.12 (m, 12H), 1.44 (s, 3H, CH₃), 2.29–2.45 (s, 1H), 2.61–2.69 (m, 1H), 3.52 (td, 1H, *J* = 9.8 Hz, 6.0 Hz), 3.76 (t, 2H, *J* = 5.7 Hz, CH₂OH), 4.05 (td, 1H, *J* = 9.9, 5.5 Hz), 4.80 (d, 1H, *J* = 3.4 Hz, C₁₀-H), 5.40 (s, 1H, C₁₂-H). FABMS (*m*/*z*): 343 [M + H]⁺, 310 [M⁺ – O₂], 267 [M⁺ – O(CH₂)₃OH]. Anal. Calcd for C₁₈H₃₀O₆: C, 63.14; H, 8.83. Found: C, 62.94; H, 8.81.

7c. White solid; mp 117–118 °C; FT-IR (KBr, cm⁻¹) 3442.9, 2925.1, 2859.3, 1458.7, 1374.7, 1023.3, 761.0; ¹H NMR (300 MHz, CDCl₃) δ 0.90 (d, 3H, *J* = 7.2 Hz, CH₃), 0.94 (d, 3H, *J* = 6.0 Hz, CH₃), 1.22–2.05 (m, 14H), 1.43 (s, 3H, CH₃), 2.31–2.37 (m, 1H), 2.61–2.63 (m, 1H), 3.37–3.49 (m, 1H), 3.66 (t, 2H, *J* = 5.4 Hz, CH₂OH), 3.85–3.90 (m, 1H), 4.78 (d, 1H, *J* = 3.3 Hz, C₁₀-H), 5.39 (s, 1H, C₁₂-H); ¹³C NMR (75 MHz, CDCl₃) δ 13.22 (CH₃), 20.59 (CH₃), 24.70 (CH₂), 24.89 (CH₂), 26.39 (CH₂), 26.41 (CH₃), 29.96

(CH₂), 31.14 (CH), 34.85 (CH₂), 36.65 (CH₂), 37.70 (CH), 44.65 (CH), 52.79 (CH), 62.91 (CH₂), 68.52 (CH₂), 81.36 (C), 88.14 (CH), 102.24 (CH), 104.34 (C). FABMS (m/z): 357 [M + H]⁺, 324 [M⁺ - O₂], 267 [M⁺ - O(CH₂)₄OH]. Anal. Calcd for C₁₉H₃₂O₆: C, 64.02; H, 9.05. Found: C, 64.19; H, 9.07.

7d. White solid; mp 64–67 °C; FT-IR (KBr, cm⁻¹) 3432.5, 2932.3, 1594.3, 1459.7, 1381.8, 1351.3, 1103.3, 1026.3, 761.1; ¹H NMR (300 MHz, CDCl₃) δ 0.83 (d, 3H, *J* = 7.3 Hz, CH₃), 0.88 (d, 3H, *J* = 5.9 Hz, CH₃), 1.15–1.98 (m, 16H), 1.36 (s, 3H, CH₃), 2.29–2.38 (m, 1H), 2.53–2.54 (m, 1H), 3.32 (td, 1H, *J* = 9.7 Hz, 6.0 Hz), 3.55 (t, 2H, *J* = 6.3 Hz, <u>CH₂OH</u>), 3.76 (td, 1H, *J* = 9.7, 6.2 Hz), 4.69 (d, 1H, *J* = 3.4 Hz, C₁₀-H), 5.32 (s, 1H, C₁₂-H); ¹³C NMR (75 MHz, CDCl₃) δ 11.67 (CH₃), 19.03 (CH₃), 21.11 (CH₂), 23.12 (CH₂), 23.34 (CH₂), 24.80 (CH₃), 28.10 (CH₂), 29.59 (CH), 30.99 (CH₂), 33.32 (CH₂), 35.10 (CH₂), 36.11 (CH), 43.14 (CH), 51.24 (CH), 61.25 (CH₂), 66.97 (CH₂), 79.79 (C), 86.55 (CH), 100.63 (C), 102.74 (C). ES-MS (*m*/*z*): 393 [M + Na]⁺, 371 [M + H]⁺, 267 [M⁺ – O(CH₂)₅OH]. Anal. Calcd for C₂₀H₃₄O₆: C, 64.84; H, 9.25. Found: C, 64.73; H, 9.00.

8d. White solid; mp 90–92 °C; FT-IR (KBr, cm⁻¹) 3427.8, 2926.2, 2856.9, 1716.5, 1460.3, 1376.8, 1217.3, 1157.8, 1019.6, 762.0; ¹H NMR (200 MHz, CDCl₃) δ 0.88 (d, 3H, *J* = 7.1 Hz, CH₃), 0.95 (d, 3H, *J* = 5.5 Hz, CH₃), 1.25–2.09 (m, 16H), 1.43 (s, 3H, CH₃), 2.30–2.45 (m, 2H), 3.37–3.48 (m, 1H), 3.65 (t, 2H, *J* = 6.2 Hz, CH₂OH), 3.91 (m, 1H), 4.41 (d, 1H, *J* = 9.2 Hz, C₁₀-H), 5.33 (s, 1H, C₁₂-H). FABMS (*m*/*z*): 371 [M + H]⁺, 267 [M⁺ – O(CH₂)₅OH]. Anal. Calcd for C₂₀H₃₄O₆: C, 64.84; H, 9.25. Found: C, 64.73; H, 9.10.

7e. Oil; FT-IR (neat, cm⁻¹) 3422.5, 2930.0, 2864.6, 1596.9, 1458.5, 1353.1, 1102.7, 1016.2, 760.9; ¹H NMR (200 MHz, CDCl₃) δ 0.89 (d, 3H, *J* = 7.3 Hz, CH₃), 0.95 (d, 3H, *J* = 5.9 Hz, CH₃), 1.21–2.07 (m, 18H), 1.44 (s, 3H, CH₃), 2.29–2.38 (m, 1H), 2.59–2.63 (m, 1H), 3.37 (td, 1H, *J* = 9.6 Hz, 6.3 Hz), 3.64 (t, 2H, *J* = 6.4 Hz, <u>CH₂OH), 3.83 (td, 1H, *J* = 9.6, 6.4 Hz), 4.77 (d, 1H, *J* = 3.2 Hz, C₁₀-H), 5.39 (s, 1H, C₁₂-H); ¹³C NMR (75 MHz, CDCl₃) δ 11.73 (CH₃), 19.08 (CH₃), 23.19 (CH₂), 23.41 (CH₂), 24.16 (CH₂), 24.77 (CH₂), 24.92 (CH₃), 28.34 (CH₂), 29.66 (CH), 31.44 (CH₂), 33.40 (CH₂), 35.18 (CH₂), 36.21 (CH), 43.22 (CH), 51.33 (CH), 61.64 (CH₂), 67.02 (CH₂), 79.87 (C), 86.63 (CH), 100.68 (CH), 102.78 (C). FABMS (*m/z*): 385 [M + H]⁺, 267 [M⁺ – O(CH₂)₆OH]. Anal. Calcd for C₂₁H₃₆O₆: C, 65.60; H, 9.44. Found: C, 65.55; H, 9.41.</u>

7f. Oil; FT-IR (neat, cm⁻¹) 3461.5, 2930.8, 1592.5, 1383.3, 1352.3, 1104.0, 1025.6, 766.2; ¹H NMR (300 MHz, CDCl₃) δ 0.91 (d, 3H, *J* = 7.3 Hz, CH₃), 0.96 (d, 3H, *J* = 6.1 Hz, CH₃), 1.29–2.06 (m, 22H), 1.45 (s, 3H, CH₃), 2.32–2.38 (m, 1H), 2.61–2.64 (m, 1H), 3.36 (td, 1H, *J* = 9.5 Hz, 6.5 Hz), 3.65 (t, 2H, *J* = 6.5 Hz, <u>CH</u>₂OH), 3.83 (td, 1H, *J* = 9.5 Hz, 6.5 Hz), 4.78 (d, 1H, *J* = 3.2 Hz, C₁₀-H) 5.40 (s, 1H, C₁₂-H); ¹³C NMR (75 MHz, CDCl₃) δ 13.19 (CH₃), 20.53 (CH₃), 24.62 (CH₂), 24.86 (CH₂), 25.86 (CH₂), 26.10 (CH₂), 26.13 (CH₃), 29.45 (CH₂), 29.55 (CH₂), 37.65 (CH), 44.68 (CH), 52.78 (CH), 63.01 (CH₂), 68.58 (CH₂), 81.32 (C), 88.07 (CH), 102.10 (CH), 104.23 (C). FABMS (*m*/*z*): 413 [M + H]⁺, 267 [M⁺ – O(CH₂)₈OH]. Anal. Calcd for (C₂₃H₄₀O₆): C 66.96; H, 9.77. Found: C, 66.76; H, 9.46.

8f. Oil; FT-IR (neat, cm⁻¹) 3429.1, 2930.8, 2817.5, 1596.3, 1383.3, 1352.3, 1025.3, 766.2; ¹H NMR (200 MHz, CDCl₃) δ 0.88 (d, 3H, *J* = 7.1 Hz, CH₃), 0.95 (d, 3H, *J* = 5.6 Hz, CH₃), 1.26–2.05 (m, 22H), 1.44 (s, 3H, CH₃), 2.32–2.44 (m, 2H), 3.37–3.42 (m, 1H), 3.63 (t, 2H, *J* = 6.4 Hz, <u>CH₂OH</u>), 3.96 (m, 1H), 4.41 (d, 1H, *J* = 9.2 Hz, C₁₀-H), 5.33 (s, 1H, C₁₂-H). FABMS (*m*/*z*): 413 [M + H]⁺, 267 [M⁺ – O(CH₂)₈OH]. Anal. Calcd for (C₂₃H₄₀O₆): C, 66.96; H, 9.77. Found: C, 66.66; H, 9.56.

7g + **8g.** Oil; FT-IR (neat, cm⁻¹) 3434.4, 2929.1, 2860.1, 1456.7, 1376.3, 1230.3, 1195.5, 1102.8, 1026.0, 755.4; ¹H NMR (200 MHz, CDCl₃) δ 0.88 and 0.89 (2 × d, 3H, *J* = 6.6 and 7.3 Hz, respectively, CH₃), 0.95 (d, 3H, *J* = 5.3 Hz, CH₃), 1.20–2.06 (m, 24H), 1.43 (s, 3H, CH₃), 2.29–2.37 (m, 1H), 2.58–2.62 (m, 1H), 3.35 (td, 1H, *J* = 9.5 Hz, 6.3 Hz), 3.62 and 4.05 (2 × t, 2H, *J* = 6.4 and 6.7 Hz, respectively, together integrating for 2H), 3.82 (td, 1H, *J* = 9.6, 6.4 Hz), 4.41 and 4.76 (2 × d, 1H, *J* = 9.2 and 3.3 Hz, respectively, together integrating for 1 C₁₀-H), 5.33 and 5.38 (2 × s, 1H, together

integrating for 1 C_{12} -H); ¹³C NMR (50 MHz, CDCl₃) δ 12.97 and 13.38 (2 × CH₃), 20.73 and 21.35 (2 × CH₃), 24.82 and 25.06 (2 × CH₂), 26.10 (CH₂), 26.37 and 26.55 (2 × CH₃), 29.60 and 29.71 (CH₂), 29.89 and 29.98 (CH₂), 31.31 (CH), 33.11 (CH₂), 34.63 and 35.06 (CH₂), 36.82 (CH₂), 37.84 (CH), 44.88 and 45.72 (2 × CH), 52.05 and 52.97 (2 × CH), 63.27 and 65.01 (2 × CH₂), 68.77 and 69.53 (2 × CH₂), 80.72 and 81.53 (2 × C), 88.27 and 91.54 (2 × CH), 100.47 and 102.30 (2 × CH), 104.43 and 104.61 (2 × C). FABMS (*m*/*z*): 427 [M + H]⁺, 267 [M⁺ - O(CH₂)₉OH]. ESMS (*m*/*z*): 449 [M + Na]⁺. Anal. Calcd for $C_{24}H_{42}O_6$: C, 67.57; H, 9.92. Found: C, 67.26; H, 9.82.

7h. Oil; FT-IR (neat, cm⁻¹) 3429.5, 2927.9, 1596.4, 1459.8, 1380.8, 1351.9, 1157.3, 1025.6, 760.3; ¹H NMR (300 MHz, CDCl₃) δ 0.89 (d, 3H, *J* = 7.5 Hz, CH₃), 0.94 (d, 3H, *J* = 6.3 Hz, CH₃), 1.21–2.06 (m, 26H), 1.43 (s, 3H, CH₃), 2.31–2.36 (m, 1H), 2.59–2.63 (m, 1H), 3.35 (td, 1H, *J* = 9.6, 6.6 Hz), 3.62 (t, 2H, *J* = 6.6 Hz, <u>CH₂OH</u>), 3.81 (td, 1H, *J* = 9.6, 6.6 Hz), 4.76 (d, 1H, *J* = 3.3 Hz, C₁₀-H), 5.38 (s, 1H, C₁₂-H); ¹³C NMR (50 MHz, CDCl₃) δ 12.99 (CH₃), 20.66 (CH₃), 22.59 (CH₂), 25.09 (CH₂), 26.12 (CH₂), 26.59 (CH₃), 29.76 (CH₂), 29.89 (CH₂), 31.34 (CH), 33.01 (CH₂), 33.16 (CH₂), 34.66 (CH₂), 36.74 (CH₂), 31.52 (C), 88.29 (CH), 102.32 (CH), 104.62 (C). FABMS (*m*/*z*): 441 [M + H]⁺, 267 [M⁺ - O(CH₂)₁₀OH]. Anal. Calcd for C₂₅H₄₄O₆: C, 68.15; H, 10.07. Found: C, 68.24; H, 10.24.

7i. Oil; FT-IR (neat, cm⁻¹) 3420.6, 2928.4, 1595.5, 1382.4, 1351.8, 1104.0, 1026.9, 763.5; ¹H NMR (300 MHz, CDCl₃) δ 0.90 (d, 3H, *J* = 7.3 Hz, CH₃), 0.95 (d, 3H, *J* = 6.1 Hz, CH₃), 1.22–2.05 (m, 30H), 1.43 (s, 3H, CH₃), 2.31–2.37 (m, 1H), 2.60–2.61 (m, 1H), 3.36 (td, 1H, *J* = 9.6, 6.4 Hz), 3.63 (t, 2H, *J* = 6.5 Hz, <u>CH₂OH</u>), 3.82 (td, 1H, *J* = 9.6, 6.6 Hz), 4.77 (d, 1H, *J* = 3.3 Hz, C₁₀-H), 5.39 (s, 1H, C₁₂-H); ¹³C NMR (75 MHz, CDCl₃) δ 11.72 (CH₃), 19.07 (CH₃), 23.16 (CH₂), 23.40 (CH₂), 24.45 (CH₂), 24.91 (CH₃), 24.93 (CH₂), 28.02 (CH₂), 28.14 (CH₂), 28.25 (CH₂), 28.28 (CH₂), 28.29 (CH₂), 28.32 (CH₂), 28.35 (CH₂), 29.67 (CH), 31.50 (CH₂), 33.41 (CH₂), 35.18 (CH₂), 79.87 (C), 86.61 (CH), 100.64 (CH), 102.75 (C). ESMS (*m*/*z*): 507 [M + K]⁺, 469 [M + H]⁺, 267 [M⁺ – O(CH₂)₁₂OH]. Anal. Calcd for C₂₇H₄₈O₆: C, 69.19; H, 10.32. Found: C, 69.40; H, 10.67.

10a. White solid; mp 94–95 °C; FT-IR (KBr, cm⁻¹) 3411.0, 2930.4, 2876.7, 1455.3, 1377.8, 1220.2, 1128.9, 1023.4, 757.2; ¹H NMR (300 MHz, CDCl₃) δ 0.92 (d, 3H, *J* = 7.3 Hz, CH₃), 0.96 (d, 3H, *J* = 6.1 Hz, CH₃), 1.20–2.07 (m, 10H), 1.44 (s, 3H, CH₃), 2.32–2.38 (m, 1H), 2.58–2.68 (m, 1H), 3.59–3.77 (m, 7H), 3.90–3.98 (m, 1H), 4.84 (d, 1H, *J* = 3.3 Hz, C₁₀-H), 5.50 (s, 1H, C₁₂-H); ¹³C NMR (75 MHz, CDCl₃) δ 13.18 (CH₃), 20.58 (CH₃), 24.67 (CH₂), 24.89 (CH₂), 26.31 (CH₃), 31.12 (CH), 34.83 (CH₂), 36.60 (CH₂), 70.89 (CH₂), 72.47 (CH₂), 81.35 (C), 88.11 (CH), 102.46 (CH), 104.40 (C). FABMS (*m*/*z*): 373 [M + H]⁺, 395 [M + Na]⁺, 267 [M⁺ – O-(CH₂CH₂O)₂H]. Anal. Calcd for C₁₉H₃₂O₇: C, 61.27; H, 8.66. Found: C, 61.62; H, 8.83.

10b. Oil; FT-IR (neat, cm⁻¹) 3432.7, 2928.0, 2876.2, 1454.6, 1454.6, 1379.0, 1218.1, 1104.5, 1026.7, 985.7, 757.0; ¹H NMR (300 MHz, CDCl₃) δ 0.87 (d, 3H, *J* = 7.3 Hz, CH₃), 0.93 (d, 3H, *J* = 6.2 Hz, CH₃), 1.20–1.84 (m, 10H), 1.41 (s, 3H, CH₃), 2.34–2.35 (m, 1H), 2.59–2.65 (m, 1H), 3.57–3.72 (m, 11H), 3.91–3.94 (m, 1H), 4.80 (d, 1H, *J* = 3.3 Hz, C₁₀-H), 5.43 (s, 1H, C₁₂-H); ¹³C NMR (75 MHz, CDCl₃) δ 11.66 (CH₃), 19.06 (CH₃), 23.11 (CH₂), 23.40 (CH₂), 24.82 (CH₃), 29.56 (CH), 33.38 (CH₂), 35.12 (CH₂), 36.12 (CH), 43.16 (CH), 51.26 (CH), 60.42 (CH₂), 66.04 (CH₂), 69.27 (CH₂), 71.30 (CH₂), 79.82 (C), 86.58 (CH), 100.77 (CH), 102.77 (C). FABMS (*m*/*z*): 417 [M + H]⁺, 267 [M⁺ – (CH₂CH₂O)₃H]. ESMS (*m*/*z*): 439 [M + Na]⁺. Anal. Calcd for C₂₁H₃₆O₈: C, 60.56; H, 8.71. Found: C, 60.31; H, 8.70.

10c. Oil; FT-IR (neat, cm⁻¹) 3422.5, 2927.1, 1457.7, 1378.3, 1218.8, 1106.0, 1025.8, 985.7, 763.7; ¹H NMR (200 MHz, CDCl₃) δ 0.91 (d, 3H, *J* = 7.3 Hz, CH₃), 0.95 (d, 3H, *J* = 6.1 Hz, CH₃), 1.20–2.05 (m, 10H), 1.43 (s, 3H, CH₃), 2.36–2.37 (m, 1H), 2.60–2.62 (m, 1H), 3.64–3.74 (m, 15H), 3.91–3.96 (m, 1H), 4.83 (d, 1H, *J* = 3.3 Hz, Cl₀-H), 5.43 (s, 1H, Cl₂-H); ¹³C NMR (75 MHz, CDCl₃) δ 13.10 (CH₃),

20.51 (CH₃), 24.58 (CH₂), 24.87 (CH₂), 26.26 (CH₃), 31.01 (CH), 34.84 (CH₂), 36.59 (CH₂), 37.61 (CH), 44.62 (CH), 52.73 (CH), 61.67 (CH₂), 67.54 (CH₂), 70.39 (CH₂), 70.59 (CH₂), 70.72 (CH₂), 72.71 (CH₂), 81.27 (C), 88.03 (CH), 102.22 (CH), 104.24 (C). FABMS (m/z): 461 [M + H]⁺, 267 [M⁺ - (CH₂CH₂O)₄H]. Anal. Calcd for C₂₃H₄₀O₉: C, 59.98; H, 8.75. Found: C, 59.80; H, 8.89.

11c. Oil; FT-IR (neat, cm⁻¹) 3427.0, 2925.7, 1459.3, 1375.5, 1219.7, 1105.2, 1026.5, 769.2; ¹H NMR (200 MHz, CDCl₃) δ 0.89 (d, 3H, *J* = 7.0 Hz, CH₃), 0.98 (d, 3H, *J* = 6.7 Hz, CH₃), 1.18–2.12 (m, 10H), 1.43 (s, 3H, CH₃), 2.29–2.43 (m, 2H), 3.58–3.75(m, 15H), 4.01–4.10 (m, 1H), 4.50 (d, 1H, *J* = 9.2 Hz, C₁₀-H), 5.33 (s, 1H, C₁₂-H). FABMS (*m*/*z*): 483 [M + Na]⁺. Anal. Calcd for C₂₃H₄₀O₉: C, 59.98; H, 8.75. Found: C, 59.80; H, 8.89.

General Procedure for Formation of Hemisuccinates 12a-i and 14a-c (Compound 12a as Representative). To a solution of hydroxy ether 7a (0.200 g, 0.6 mmol) and succinic anhydride (0.300 g, 3.0 mmol) in CH2Cl2 (30 mL) was added pyridine (0.24 mL, 3.0 mmol). The reaction mixture was stirred at the room temperature for 15 h. Then the reaction mixture was quenched with 10% aqueous HCl solution (20 mL) and extracted with ether (3 \times 20 mL). The combined organic layer was washed with water (20 mL) and then with brine (25 mL), dried over anhydrous Na2SO4, and concentrated under reduced pressure. The crude product on column chromatography over silica gel using ethyl acetate/hexane (1:4) as eluant furnished pure 12a as an oil (0.160 g, 62% yield). FT-IR (neat,cm⁻¹) 2927.5, 2819.8, 1731.5, 1452.5, 1379.2, 1362.3, 1211.4, 1173.4, 1102.5, 1025.9, 760.5; ¹H NMR (200 MHz, CDCl₃) δ 0.90 (d, 3H, J = 7.3 Hz, CH₃), 0.95 (d, $3H_{1}J = 5.8 Hz_{1} CH_{3}$, 1.10-2.12 (m, 10H), $1.43 (s, 3H, CH_{3})$, 2.31-2.41 (m, 1H), 2.66 (m, 5H), 3.59–3.76 (m, 1H), 3.65 and 3.98 (2 \times m, 1H, OCH₂CH₂OCO(CH₂)₂COOH), 3.94-4.05 (m, 1H), 4.26 (m, 1H), 4.81 (d, 1H, J = 3.3 Hz, C_{10} -H), 5.43 (s, 1H, C_{12} -H). FABMS (m/z): 435 $[M + Li]^+$, 397 $[(M^+ - O_2) + H]^+$, 267 $[M^+ - O_2)$ $OCH_2CH_2OCO(CH_2)_2COOH$]. 12a on treatment with CH_2N_2 in ether and subsequent purification by column chromatography furnished ester 13a as a white solid. Mp 70-71 °C; FT-IR (KBr, cm⁻¹) 1725.0; ¹H NMR (200 MHz, CDCl₃) δ 0.83 (d, 3H, J = 7.3 Hz, CH₃), 0.88 (d, 3H, J = 5.8 Hz, CH₃), 1.14–1.99 (m, 10H), 1.36 (s, 3H, CH₃), 2.22-2.36 (m, 1H), 2.57-2.66 (m, 5H), 3.52-3.69 (m, 1H), 3.62 (s, 3H, O<u>CH</u>₃), 3.87–3.98 (m, 1H), 4.16–4.25 (t, 2H, J = 4.9 Hz, $OCH_2CH_2OCO(CH_2)_2COOCH_3)$, 4.74 (d, 1H, J = 3.2 Hz, C_{10} -H), 5.35 (s, 1H, C₁₂-H). FABMS (m/z): 443 [M + H]⁺, 267 [M⁺ -OCH₂CH₂OCO(CH₂)₂COOCH₃]. Anal. Calcd for C₂₂H₃₄O₉: C, 59.71; H, 7.74. Found: C, 60.12; H, 7.85.

Hemisuccinates 12b-i and 14a-c were prepared using the above procedure.

12b. Yield 78%; oil; FT-IR (neat, cm^{-1}) 2923.6, 2845.3, 1732.0, 1451.7, 1380.0, 1360.5, 1210.7, 1172.4, 1102.1, 1027.1, 760.3; ¹H NMR (400 MHz, CDCl₃) δ 0.90 (d, 3H, J = 7.4 Hz, CH₃), 0.96 (d, 3H, J = 6.0 Hz, CH₃), 1.20-2.07 (m, 12H), 1.44 (s, 3H, CH₃), 2.33-2.43 (m, 1H), 2.77 (m, 5H), 3.41 (td, 1H, J = 10.5, 6.0 Hz), 3.94 (td, 1H, J = 10.5, 6.0 Hz), 4.15 (m, 1H), 4.27 (m, 1H), 4.88 (d, 1H, J = 3.6 Hz, C₁₀-H), 5.44 (s, 1H, C₁₂-H). FABMS (m/z): 449 [M + Li]⁺, 267 $[M^+ - O(CH_2)_3 OCO(CH_2)_2 COOH]$. 12b on treatment with CH_2N_2 in ether and subsequent purification by column chromatography furnished ester 13b as an oil. FT-IR (neat, cm⁻¹) 1734.0; ¹H NMR $(200 \text{ MHz}, \text{CDCl}_3) \delta 0.89 \text{ (d, 3H, } J = 7.4 \text{ Hz}, \text{CH}_3\text{)}, 0.95 \text{ (d, 3H, } J =$ 5.6 Hz, CH₃), 1.25-2.06 (m, 12H), 1.43 (s, 3H, CH₃), 2.31-2.43 (s, 1H), 2.63 (m, 5H), 3.38-3.96 (m, 2H), 3.64 (s, 3H, OCH₃), 4.17 (t, 2H, J = 6.2 Hz, <u>CH</u>₂OCO(CH₂)₂COOCH₃), 4.77 (d, 1H, J = 3.3 Hz, C₁₀-H), 5.40 (s, 1H, C₁₂-H). ESMS (m/z): 479 [M + Na]⁺, 267 [M⁺ - $O(CH_2)_3OCO(CH_2)_2COOH$]. Anal. Calcd for $C_{23}H_{36}O_9$: C, 60.51; H, 7.95. Found: C, 60.85; H, 7.90.

12c. Yield 68%; oil; FT-IR (neat, cm⁻¹) 2926.8, 2875.1, 1732.4, 1450.1, 1379.2, 1362.2, 1215.8, 1173.3, 1102.8, 1027.3, 769.8; ¹H NMR (200 MHz, CDCl₃) δ 0.90 (d, 3H, *J* = 7.3 Hz, CH₃), 0.95 (d, 3H, *J* = 5.9 Hz, CH₃), 1.21–2.07 (m, 14H), 1.44 (s, 3H, CH₃), 2.29–2.43 (s, 1H), 2.65 (m, 5H), 3.38–3.46 (m, 1H), 3.80–3.88 (m, 1H), 4.12 (brs, 2H, <u>CH₂OCO(CH₂)₂COOH</u>), 4.80 (d, 1H, *J* = 2.8 Hz, C₁₀-H), 5.42 (s, 1H, C₁₂-H); ¹³C NMR (75 MHz, CDCl₃) δ 11.73 (CH₃), 19.06 (CH₃), 23.16 (CH₂), 23.35 (CH₂), 24.23 (CH₂), 24.80 (CH₃),

24.97 (CH₂), 27.69 (CH₂), 28.39 (CH₂), 29.59 (CH), 33.34 (CH₂), 35.11 (CH₂), 36.10 (CH), 43.13 (CH), 51.29 (CH), 63.45 (CH₂), 66.67 (CH₂), 79.85 (C), 86.69 (CH), 100.72 (CH), 102.93 (C), 171.02 (C), 175.95 (C). FABMS (*m*/*z*): 457 [M + H] ⁺, 267 [M⁺ – O(CH₂)₄OCO(CH₂)₂COOH]. **12c** on treatment with CH₂N₂ in ether and subsequent purification by column chromatography furnished ester **13c** as an oil. FT-IR (neat, cm⁻¹) 2923.9, 2873.7, 1737.7, 1436.9, 1361.7, 1161.1, 1103.2, 1029.9, 760.1; ¹H NMR (400 MHz, CDCl₃) δ 0.91 (d, 3H, *J* = 7.3 Hz, CH₃), 0.96 (d, 3H, *J* = 6.0 Hz, CH₃), 1.21–2.07 (m, 14H), 1.43 (s, 3H, CH₃), 2.33–2.41 (m, 1H), 2.64 (m, 5H), 3.38 (td, 1H, *J* = 10.1, 5.7 Hz), 3.68 (s, 3H, O<u>CH₃</u>), 3.87 (td, 1H, *J* = 10.1, 5.7 Hz), 4.12 (t, 2H, *J* = 6.0 Hz, CH₂OCO(CH₂)₂COOCH₃), 4.78 (d, 1H, *J* = 3.6 Hz, Cl₀-H), 5.38 (s, 1H, Cl₁²-H). FABMS (*m*/*z*): 471 [M + H] ⁺. Anal. Calcd for C₂₄H₃₈O₉: C, 61.26; H, 8.14. Found: C, 60.81; H, 8.03.

12d. Yield 84%; oil; FT-IR (neat, cm⁻¹) 2931.4, 2373.4, 1732.0, 1591.8, 1351.4, 1164.1, 1105.0, 1027.2, 756.0; ¹H NMR (200 MHz, $CDCl_3$) δ 0.91 (d, 3H, J = 7.3 Hz, CH₃), 0.96 (d, 3H, J = 5.8 Hz, CH₃), 1.23-2.06 (m, 16H), 1.43 (s, 3H, CH₃), 2.36 (m, 1H), 2.60-2.69 (m, 5H), 3.39 (td, 1H, J = 9.7, 6.2 Hz), 3.83 (td, 1H, J = 9.7, 6.2 Hz), 4.12 (t, 2H, J = 6.4 Hz, CH₂OCO(CH₂)₂COOH), 4.78 (d, 1H, $J = 3.4 \text{ Hz}, C_{10}\text{-H}), 5.41 (s, 1H, C_{12}\text{-H}); {}^{13}\text{C} \text{ NMR} (75 \text{ MHz}, \text{CDCl}_3)$ δ 11.74 (CH₃), 19.07 (CH₃), 21.40 (CH₂), 23.21 (CH₂), 23.42 (CH₂), 24.88 (CH₃), 26.99 (CH₂), 27.68 (CH₂), 27.90 (CH₂), 28.41 (CH₂), 29.66 (CH), 33.41 (CH₂), 35.13 (CH₂), 36.21 (CH), 43.23 (CH), 51.34 CH), 63.46 (CH₂), 66.95 (CH₂), 79.92 (C), 86.67 (CH), 100.66 (CH), 102.89 (C), 170.96 (C), 175.78 (C). ESMS (m/z): 493 $[M + Na]^+$, 488 $[M + NH_4]^+$, 509 $[M + K]^+$. 12d on treatment with CH₂N₂ in ether and subsequent purification by column chromatography furnished ester 13d as an oil. FT-IR (neat, cm⁻¹) 1737.0; ¹H NMR (400 MHz, CDCl₃) δ 0.89 (d, 3H, J = 7.3 Hz, CH₃), 0.95 (d, $3H_1 J = 5.7 Hz_1 CH_3$, 1.21-2.11 (m, 16H), $1.43 (s, 3H, CH_3)$, 2.95-2.45 (m, 1H), 2.62 (m, 5H), 3.37 (td, 1H, J = 9.5, 6.5 Hz), 3.69 (s, 3H, OCH_3), 3.84 (td, 1H, J = 9.6, 6.5 Hz), 4.09 (t, 2H, J = 6.4 Hz, CH₂CH₂OCO(CH₂)₂COOCH₃), 4.77 (d, 1H, J = 3.0 Hz, C₁₀-H), 5.38 (s, 1H, C_{12} -H). FABMS (m/z): 507 [M + Na]⁺, 523 [M + K]⁺. Anal. Calcd for C25H40O9: C, 61.96; H, 8.32. Found: C, 62.30; H, 8.63.

12e. Yield 71%; oil; FT-IR (neat, cm⁻¹) 2935.5, 2869.9, 1735.8, 1442.7, 1367.4, 1218.9, 1164.9, 1103.2, 1018.3, 758.0; ¹H NMR (200 MHz, CDCl₃) δ 0.90 (d, 3H, J = 7.3 Hz, CH₃), 0.94 (d, 3H, J = 5.7 Hz, CH₃), 1.24–2.11 (m, 18H), 1.43 (s, 3H, CH₃), 2.31–2.43 (m, 1H), 2.62-2.69 (m, 5H), 3.35-3.41 (m, 1H), 3.76-3.87 (m, 1H), 4.12 (t, 2H, J = 6.0 Hz, CH₂OCO(CH₂)₂COOH), 4.80 (d, 1H, J = 3.2 Hz, C₁₀-H), 5.42 (s, 1H, $\overline{C_{12}}$ -H); ¹³C NMR (75 MHz, CDCl₃) δ 11.73 (CH₃), 19.11 (CH₃), 23.17 (CH₂), 23.35 (CH₂), 24.23 (CH₂), 24.45 (CH₂), 24.83 (CH₃), 27.27 (CH₂), 27.77 (CH₂), 28.12 (CH₂), 28.40 (CH₂), 29.65 (CH), 33.42 (CH₂), 35.36 (CH₂), 36.17 (CH), 43.31 (CH), 51.46 (CH), 63.91 (CH₂), 67.06 (CH₂), 80.12 (C), 86.87 (CH), 100.82 (CH), 103.07 (C), 171.48 (C), 178.24 (C); ESMS (m/z): 507 $[M + Na]^+$, 267 $[M^+ - O(CH_2)_6 OCO(CH_2)_2 COOH]^+$. 12e on treatment with CH_2N_2 in ether and subsequent purification by column chromatography furnished ester 13e as an oil. FT-IR (neat, cm⁻¹) 2931.6, 2746.4, 1695.3, 1419.5, 1311.5, 1203.5, 920.0, 771.5; ¹H NMR (300 MHz, CDCl₃) δ 0.90 (d, 3H, J = 7.2 Hz, CH₃), 0.95 (d, 3H, J = 6.0 Hz, CH₃), 1.23–2.06 (m, 18H), 1.44 (s, 3H, CH₃), 2.37– 2.38 (m, 1H), 2.61-2.64 (m, 5H), 3.35-3.38 (m, 1H), 3.70 (s, 3H, OCH_3), 3.82–3.85 (m, 1H), 4.09 (t, 2H, J = 6.6 Hz, CH_2OCO - $(CH_2)_2COOCH_3)$, 4.78 (d, 1H, J = 3.0 Hz, C_{10} -H), 5.39 (s, 1H, C_{12} -H). FABMS (m/z): 499 $[M + H]^+$, 521 $[M + Na]^+$. Anal. Calcd for C26H42O9: C, 62.63; H, 8.49. Found: C, 62.99; H, 8.77.

12f. Yield 70%; oil; FT-IR (neat, cm⁻¹) 2920.8, 2853.8, 2363.6, 1726.0, 1594.1, 1382.0, 1351.8, 1218.9, 1164.5, 1103.7, 1027.6, 761.4; ¹H NMR (200 MHz, CDCl₃) δ 0.91 (d, 3H, *J* = 7.3 Hz, CH₃), 0.97 (d, 3H, *J* = 5.8 Hz, CH₃), 1.27–2.12 (m, 22H), 1.45 (s, 3H, CH₃), 2.33–2.46 (m, 1H), 2.59–2.69 (m, 5H), 3.37–3.40 (m, 1H), 3.79–3.85 (m, 1H), 4.11 (t, 2H, *J* = 6.6 Hz, <u>CH₂OCO(CH₂)₂COOH), 4.79 (d, 1H, *J* = 3.2 Hz, C₁₀-H), 5.41 (s, 1H, C₁₂-H); ¹³C NMR (75 MHz, CDCl₃) δ 11.72 (CH₃), 19.08 (CH₃), 23.18 (CH₂), 23.42 (CH₂), 24.47 (CH₂), 24.77 (CH₂), 24.88 (CH₃), 27.25 (CH₂), 27.69 (CH₂), 27.83 (CH₂), 28.29 (CH₂), 28.42 (CH₂), 29.65 (CH), 33.41 (CH₂), 35.19</u>

(CH₂), 36.23 (CH), 43.20 (CH), 51.33 (CH), 63.67 (CH₂), 67.11 (CH₂), 79.88 (C), 86.68 (CH), 100.65 (CH), 102.84 (C), 170.98 (C), 177.88 (C). ESMS (*m*/*z*): 512 [M + Na] ⁺, 267 [M⁺ – O(CH₂)₈OCO(CH₂)₂COOH]⁺. **12f** on treatment with CH₂N₂ in ether and subsequent purification by column chromatography furnished ester **13f** as an oil. FT-IR (neat, cm⁻¹) 2932.4, 2300.4, 1736.4, 1448.7, 1366.3, 1162.8, 1102.3, 1028.1, 874.0; ¹H NMR (200 MHz, CDCl₃) δ 0.89 (d, 3H, *J* = 7.3 Hz, CH₃), 0.95 (d, 3H, *J* = 5.8 Hz, CH₃), 1.18–2.12 (m, 22H), 1.44 (s, 3H, CH₃), 2.31–2.46 (m, 1H), 2.63 (m, 5H), 3.35 (td, 1H, *J* = 9.9 Hz, 6.1 Hz), 3.69 (s, 3H, O<u>CH₃</u>), 3.82 (td, 1H, *J* = 9.7 Hz, 6.4 Hz), 4.08 (t, 2H, *J* = 6.6 Hz, CH₂OCO(CH₂)₂COOCH₃), 4.77 (d, 1H, *J* = 3.3 Hz, C₁₀-H), 5.39 (s, 1H, C₁₂-H). FABMS (*m*/*z*): 527 [M + H]⁺, 549 [M + Na]⁺, 267 [M⁺ – O(CH₂)₈OCO(CH₂)₂COOCH₃]⁺. Anal. Calcd for C₂₈H₄₆O₉: C, 63.86; H, 8.80. Found: C, 63.96; H, 9.14.

12g. Yield 83%; oil; FT-IR (neat, cm⁻¹) 2930.8, 2374.3, 1722.0, 1596.2, 1352.0, 1026.6, 761.6; ¹H NMR (200 MHz, CDCl₃) δ 0.88 and 0.89 (2 × d, together integrating for 3H, J = 6.8 and 7.3 Hz, CH₃), 0.95 (d, 3H, J = 5.7 Hz, CH₃), 1.25–2.04 (m, 24H), 1.43 (s, 3H, CH₃), 2.29-2.43 (m, 1H), 2.62 (m, 5H), 3.31-3.46 and 3.62-3.67 $(2 \times m, \text{ together integrating for 1H}), 3.76-3.97$ $(2 \times m, \text{ together})$ integrating for 1H), 4.08 (t, 2H, J = 6.4 Hz, CH₂OCO(CH₂)₂COOH), 4.42 and 4.78 (2 \times d, 1H, J = 9.2 and 3.1 Hz, respectively, together integrating for C_{10} -H), 5.33 and 5.39 (2 × s, 1H, together integrating for C₁₂-H); ¹³C NMR (50 MHz, CDCl₃) δ 13.01 and 13.42 (2 × CH₃), 20.67 and 20.77 (2 × CH₃), 24.85 and 25.09 (2 × CH₂), 26.23 and 26.56 (2 × CH₃), 28.93 (CH₂), 29.55 (CH₂), 30.07 (CH₂), 31.34 (CH), 34.66 and 35.09 (2 × CH₂), 36.85 (CH₂), 37.87 (CH), 44.91 and 45.73 (2 \times CH), 51.01 (CH), 65.40 (CH₂), 68.80 and 69.59 (2 \times CH₂), 81.56 (C), 88.35 and 91.58 (2 × CH), 100.54 and 102.31 (2 × CH), 104.49 (C), 173.04 (C), 177.40 (C). ESMS (m/z): 527 [M + H] +, 481 [M⁺ – COOH], 267 [M⁺ – O(CH₂)₉OCO(CH₂)₂COOH]⁺. 12g on treatment with CH_2N_2 in ether and subsequent purification by column chromatography furnished ester 13g as an oil. FT-IR (neat, cm⁻¹) 1732.0; ¹H NMR (200 MHz, CDCl₃) δ 0.89 (d, 3H, J = 7.3 Hz, CH_3), 0.95 (d, 3H, J = 5.8 Hz, CH_3), 1.25–2.04 (m, 24H), 1.43 (s, 3H, CH₃), 2.29–2.43 (m, 1H), 2.62 (m, 5H), 3.23–3.41 (m, 1H), 3.69 (s, 3H, OCH₃), 3.76-3.97 (m, 1H), 4.08 (t, 2H, J = 6.5 Hz, <u>CH</u>₂OCO(CH₂)₂COOCH₃), 4.41 and 4.77 (d, 1H, J = 9.7 and 2.7 Hz, respectively, together integrating for C₁₀-H), 5.33 and 5.38 (s, 1H, together integrating for C₁₂-H). FABMS (m/z): 539 [M⁺ – H], 267 $[M^+ - O(CH_2)_9OCO(CH_2)_2COOCH_3]^+$. Anal. Calcd for $C_{29}H_{48}O_9$: C, 64.42; H, 8.95. Found: C, 64.78; H, 9.16.

12h. Yield 83%; oil; FT-IR (neat, cm⁻¹) 2928.5, 2818.9, 2181.8, 1726.0, 1595.5, 1383.0, 1351.8, 1025.9, 763.5; ¹H NMR (300 MHz, $CDCl_3$) δ 0.91 (d, 3H, J = 7.3 Hz, CH₃), 0.96 (d, 3H, J = 5.8 Hz, CH₃), 1.23-2.07 (m, 26H), 1.45 (s, 3H, CH₃), 2.31-2.41 (m, 1H), 2.63-2.67 (m, 5H), 3.36-3.39 (m, 1H), 3.81-3.85 (m, 1H), 4.10 (t, 2H, J = 6.6 Hz, <u>CH₂OCO(CH₂)₂COOH</u>), 4.79 (d, 1H, J = 3.4 Hz, C_{10} -H), 5.41 (s, 1H, C_{12} -H); ¹³C NMR (75 MHz, CDCl₃) δ 11.75 (CH₃), 19.06 (CH₃), 23.20 (CH₂), 23.40 (CH₂), 24.58 (CH₂), 24.90 (CH₃), 27.26 (CH₂), 27.69 (CH₂), 27.94 (CH₂), 28.16 (CH₂), 28.37 (CH₂), 29.68 (CH), 33.46 (CH₂), 35.20 (CH₂), 36.21 (CH), 43.25 (CH), 51.37 (CH), 63.81 (CH₂), 67.20 (CH₂), 79.93 (C), 86.69 (CH), 100.64 (CH), 102.79 (C), 171.00 (C), 175.73 (C). FABMS (m/z): 541 $[M + H]^+$, 267 $[M^+ - O(CH_2)_{10}OCO(CH_2)_2COOH]^+$. 12h on treatment with CH₂N₂ in ether and subsequent purification by column chromatography furnished ester 13h as an oil. FT-IR (neat, cm⁻¹) 2929.7, 2860.2, 1737.7, 1461.9, 1164.9, 1024.1, 756.0; ¹H NMR (200 MHz, CDCl₃) δ 0.91 (d, 3H, J = 7.3 Hz, CH₃), 0.96 (d, 3H, J = 6.0 Hz, CH₃), 1.22-2.07 (m, 26H), 1.44 (s, 3H, CH₃), 2.33-2.43 (m, 1H), 2.64 (m, 5H), 3.36 (td, 1H, J = 10.1 Hz, 6.1 Hz), 3.71 (s, 3H, OCH_3 , 3.83 (td, 1H, J = 10.1 Hz, 6.1 Hz), 4.28 (t, 2H, J = 6.5 Hz, <u>CH₂OCO(CH₂)₂COOCH₃), 4.79 (d, 1H, J = 3.2 Hz, C₁₀-H), 5.40 (s,</u> 1H, C₁₂-H). FABMS (m/z): 555 [M + H]⁺, 267 [M⁺ $O(CH_2)_{10}OCO(CH_2)_2COOCH_3]^+$. Anal. Calcd for $C_{30}H_{50}O_9$: C, 64.96; H, 9.09. Found: C, 64.77; H, 9.13.

12i. Yield 83%; oil; FT-IR (neat, cm⁻¹) 2819.8, 2375.9, 1724.0, 1595.7, 1352.2, 1017.0, 763.4; ¹H NMR (200 MHz, CDCl₃) δ 0.89 (d, 3H, *J* = 7.3 Hz, CH₃), 0.95 (d, 3H, *J* = 5.8 Hz, CH₃), 1.21–2.07 (m,

30H), 1.44 (s, 3H, CH₃), 2.31-2.43 (m, 1H), 2.61-2.67 (m, 5H), 3.37 (td, 1H, J = 9.5 Hz, 6.2 Hz), 3.80 (td, 1H, J = 9.5 Hz, 6.5 Hz), 4.09 (t, 2H, J = 6.6 Hz, <u>CH₂OCO(CH₂)₂COOH</u>), 4.78 (d, 1H, J = 3.4 Hz, C₁₀-H), 5.39 (s, 1H, $\overline{C_{12}}$ -H); ¹³C NMR (75 MHz, CDCl₃) δ 13.00 (CH₃), 20.67 (CH₃), 24.85 (CH₂), 25.09 (CH₂), 26.24 (CH₂), 26.40 (CH₂), 26.60 (CH₃), 28.94 (CH₂), 29.19 (CH₂), 29.36 (CH₂), 29.92 (CH₂), 30.07 (CH₂), 31.36 (CH), 33.01 (CH₂), 36.87 (CH₂), 37.87 (CH₂), 44.92 (CH), 53.02 (CH), 65.44 (CH₂), 68.85 (CH₂), 81.68 (C), 88.32 (CH), 100.32 (CH), 102.32 (C), 172.63 (C), 177.01 (C). ESMS (m/z): 569 $[M + H]^+$, 591 $[M + Na]^+$, 607 $[M + K]^+$, 267 $[M^+ - O(CH_2)_{12}OCO(CH_2)_2COOH]^+$. 12i on treatment with CH₂N₂ in ether and subsequent purification by column chromatography furnished ester 13i as an oil. FT-IR (neat, cm⁻¹) 1740.0; ¹H NMR $(200 \text{ MHz}, \text{CDCl}_3) \delta 0.89 \text{ (d, 3H, } J = 7.3 \text{ Hz}, \text{CH}_3\text{)}, 0.95 \text{ (d, 3H, } J =$ 5.9 Hz, CH₃), 1.20–2.07 (m, 30H), 1.44 (s, 3H, CH₃), 2.29–2.38 (m, 1H), 2.59-2.63 (m, 5H), 3.36 (td, 1H, J = 9.5 Hz, 6.5 Hz), 3.69 (s, 3H, OCH₃), 3.82 (td, 1H, J = 9.8 Hz, 6.5 Hz), 4.08 (t, 2H, J = 6.6 Hz, <u>CH₂OCO(CH₂)₂COOCH₃), 4.77 (d, 1H, J = 3.5 Hz, C₁₀-H), 5.39 (s,</u> 1H, C₁₂-H). ESMS (m/z): 583 $[M + H]^+$, 605 $[M + Na]^+$. Anal. Calcd for C32H54Oo: C, 65.95; H, 9.34. Found: C, 65.97; H, 9.44.

14a. Yield 51%; oil; FT-IR (neat, cm⁻¹) 2926.5, 1727.5, 1627.5, 1568.1, 1381.2, 1229.3, 1168.1, 1084.2, 1022.4, 620.2; ¹H NMR (200 MHz, CDCl₃) δ 0.91 (d, 3H, J = 7.6 Hz, CH₃), 0.95 (d, 3H, J = 6.0 Hz, CH₃), 1.25-2.13 (m, 10H), 1.43 (s, 3H, CH₃), 2.31-2.38 (m, 1H), 2.67 (m, 5H), 3.60-3.70 (m, 5H), 3.89-3.92 (m, 1H), 4.22-4.26 (m, 2H, $\underline{CH}_2OCO(CH_2)_2COOH$), 4.86 (d, 1H, J = 2.7 Hz, C_{10} -H), 5.45 (s, 1H, C_{12} -H); ¹³C NMR (75 MHz, CDCl₃) δ 11.64 (CH₃), 19.07 (CH₃), 23.14 (CH₂), 23.39 (CH₂), 24.78 (CH₃), 27.77 (CH₂), 30.83 (CH), 33.40 (CH₂), 35.14 (CH₂), 36.16 (CH), 43.18 (CH), 51.29 (CH), 62.67 (CH₂), 65.81 (CH₂), 67.59 (CH₂), 69.21 (CH₂), 79.89 (C), 88.62 (CH), 100.69 (CH), 102.95 (C), 172.39 (C), 175.85 (C). ESMS (m/z): 473 $[M + H]^+$, 267 $[M^+ - O(CH_2)_2O^ (CH_2)_2OCO(CH_2)_2COOH^{\dagger}$. 14a on treatment with CH_2N_2 in ether and subsequent purification by column chromatography furnished ester 15a as an oil. FT-IR (neat, cm⁻¹) 1730.0; ¹H NMR (400 MHz, CDCl₃) δ 0.91 (d, 3H, J = 7.6 Hz, CH₃), 0.94 (d, 3H, J = 5.6 Hz, CH₃), 1.21-2.07 (m, 10H), 1.44 (s, 3H, CH₃), 2.33-2.43 (m, 1H), 2.68 (m, 5H), 3.62-3.73 (m, 5H), 3.71 (s, 3H, OCH₃), 3.95 (m, 1H), 4.23 (t, 2H, J = 5.1 Hz, <u>CH₂OCO(CH₂)₂COOCH₃), 4.85 (d,</u> 1H, J = 2.8 Hz, C_{10} -H), 5.43 (s, 1H, C_{12} -H). FABMS (m/z): 487 [M + H]⁺, 267 [M⁺ – O(CH₂CH₂O)₂CO(CH₂)₂COOCH₃]⁺. Anal. Calcd for C24H38O10: C, 59.24; H, 7.87. Found: C, 59.29; H, 8.13.

14b. Yield 78%; oil; FT-IR (neat, cm⁻¹) 2929.2, 2877.5, 1733.0, 1452.9, 1380.1, 1218.6, 1167.4, 1136.1, 1107.5, 1025.8, 760.9; ¹H NMR (200 MHz, CDCl₃) δ 0.91 (d, 3H, J = 7.6 Hz, CH₃), 0.94 (d, 3H, J = 6.9 Hz, CH₃), 1.21-2.14 (m, 10H), 1.43 (s, 3H, CH₃), 2.31-2.43 (m, 1H), 2.65 (m, 5H), 3.64-3.67 (m, 9H), 3.92-3.99 (m, 1H), 4.27 (t, 2H, J = 4.5 Hz, <u>CH₂OCO(CH₂)₂COOH</u>), 4.85 (d, 1H, J = 3.1 Hz, C₁₀-H), 5.43 (s, 1H, $\overline{C_{12}}$ -H); ¹³C NMR (75 MHz, CDCl₃) δ 11.58 (CH₃), 19.00 (CH₃), 23.12 (CH₂), 23.40 (CH₂), 24.74 (CH₃), 27.19 (CH₂), 27.62 (CH₂), 27.80 (CH₂), 28.37 (CH₂), 29.56 (CH), 33.38 (CH₂), 35.11 (CH₂), 36.15 (CH), 43.14 (CH), 51.26 (CH), 62.57 (CH₂), 66.01 (CH₂), 67.70 (CH₂), 69.19 (CH₂), 79.95 (C), 86.56 (CH), 100.84 (CH), 102.80 (C), 170.98 (C), 174.90 (C). FABMS (m/z): 517 $[M + H]^+$, 267 $[M^+ - O(CH_2CH_2O)_3CO(CH_2)_2$ -COOH]⁺. 14b on treatment with CH₂N₂ in ether and subsequent purification by column chromatography furnished ester 15b as an oil. FT-IR (neat, cm⁻¹) 2873.7, 1739.7, 1438.8, 1348.1, 1105.1, 1029.9, 758.0; ¹H NMR (400 MHz, CDCl₃) δ 0.90 (d, 3H, J = 7.6 Hz, CH₃), 0.94 (d, 3H, J = 6.0 Hz, CH₃), 1.19-2.05 (m, 10H), 1.42 (s, 3H, CH₃), 2.33-2.41 (m, 1H), 2.61-2.69 (m, 5H), 3.58-3.70 (m, 9H), 3.68 (s, 3H, OCH_3), 3.92–3.97 (m, 1H), 4.25 (t, 2H, J = 5.0 Hz, <u>CH₂OCO(CH₂)₂COOCH₃), 4.84 (d, 1H, J = 3.2 Hz, C₁₀-H), 5.42</u> (s, 1H, C₁₂-H). FABMS (m/z): 531 [M + H]⁺, 537 [M + Li]⁺, 553 $[M + Na]^+$. Anal. Calcd for $C_{26}H_{42}O_{11}$ ·0.2 H_2O : C, 58.85; H, 7.98. Found: C, 58.69; H, 7.97.

14c. Yield 66%; oil; FT-IR (neat, cm⁻¹) 2925.9, 1734.1, 1454.1, 1379.3, 1218.5, 1106.0, 1027.3, 985.6, 758.2; ¹H NMR (200 MHz, CDCl₃) δ 0.90 (d, 3H, J = 7.4 Hz, CH₃), 0.94 (d, 3H, J = 6.1 Hz, CH₃), 1.20–2.04 (m, 10H), 1.43 (s, 3H, CH₃), 2.31–2.37 (m, 1H),

2.63-2.69 (m, 5H), 3.56-3.69 (m, 13H), 3.85-3.98 (m, 1H), 4.24-4.29 (m, 2H, <u>CH</u>₂OCO(CH₂)₂COOCH₃), 4.83 (d, 1H, J = 3.4 Hz, C₁₀-H), 5.42 (s, 1H, C₁₂-H); ¹³C NMR (75 MHz, CDCl₃) δ 11.68 (CH₃), 19.09 (CH₃), 23.12 (CH₂), 23.40 (CH₂), 24.83 (CH₃), 27.73 (CH₂), 27.99 (CH₂), 28.39 (CH₂), 29.56 (CH), 33.37 (CH₂), 35.12 (CH₂), 36.13 (CH), 43.14 (CH), 51.25 (CH), 62.53 (CH₂), 66.03 (CH₂), 67.70 (CH₂), 69.16 (CH₂), 69.28 (CH₂), 69.36 (CH₂), 79.84 (C), 86.59 (CH), 100.76 (CH), 102.82 (C), 170.86 (C), 174.50 (C). FABMS (m/z): 585 [M + Na]⁺, 599 [M + K]⁺, 267 [M⁺ - O- $(CH_2CH_2O)_4CO(CH_2)_2COOH^{\dagger}$. 14c on treatment with CH_2N_2 in ether and subsequent purification by column chromatography furnished ester 15c as an oil. FT-IR (neat, cm⁻¹) 2942.0, 1740.0, 1634.3, 1454.6, 1266.0, 1112.3, 1030.1, 756.2; ¹H NMR (300 MHz, CDCl₃) δ 0.91 (d, 3H, J = 7.2 Hz, CH₃), 0.94 (d, 3H, J = 6.3 Hz, CH₃), 1.21–2.05 (m, 10H), 1.43 (s, 3H, CH₃), 2.31–2.37 (m, 1H), 2.61-2.68 (m, 5H), 3.61-3.71 (m, 13H), 3.69 (s, 3H, OCH3), 3.91-3.97 (m, 1H), 4.25 (t, 2H, J = 4.8 Hz, <u>CH₂OCO(CH₂)₂COOCH₃),</u> 4.82 (d, 1H, J = 3.3 Hz, C_{10} -H), 5.42 (s, 1H, C_{12} -H). FABMS (m/z): 575 $[M + H]^+$, 267 $[M^+ - O(CH_2CH_2O)_4CO(CH_2)_2COOH]^+$. Anal. Calcd for C₂₈H₄₆O₁₂: C, 58.52; H, 8.07. Found: C, 58.12; H, 8.24.

ASSOCIATED CONTENT

Supporting Information

Table of purity data, evidence for stability of compound 12h, table of NMR data of the final products 13a-i and 15a-c, and ¹H NMR and ¹³C NMR spectra of compounds 7d, 7e, 7h, 7i, 10b, 12d, 12e, 12f, and 12h. This material is available free of charge via the Internet at http://pubs.acs.org.

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(8) (a) Compounds 7a-c and 10a have been prepared earlier: (a1) Venugopalan, B.; Karnik, P. J.; Bapat, C. P.; Chatterjee, D. K.; Iyer, N.; Lepcha, D. Antimalarial activity of new ethers and thioethers of dihydroartemisinin. *Eur. J. Med. Chem.* 1995, 30, 697–706. (a2) Galal, A. M.; Ross, S. A.; Jacob, M.; ElSohly, M. A. Antifungal activity of artemisinin derivatives. *J. Nat. Prod.* 2005, 68, 1274–1276. (b) α -Isomers 8d, 8f, and 11c were isolated and characterized but were not used for the synthesis of the hemisuccinate derivatives. Compounds 7g and 8g could not be separated and were used as a mixture for the preparation of hemisuccinate derivative 12g.

(9) (a) Peters, W. Techniques for the Study of Drug Response in Experimental Malaria. In Chemotherapy and Drug Resistance in Malaria; Academic Press: London, 1970; pp 64-136. (b) For the in vivo antimalarial efficacy test, the blood schizontocidal activity of the test compounds was evaluated in rodent model using multidrug-resistant strain of Plasmodium yoelii nigeriensis. The colony bred Swiss mice of either sex (20 \pm 2 g) were inoculated intraperitoneally with 1 \times 10⁵ P. yoelii (MDR) parasites on day 0, and treatment was administered to a group of five or six mice at each dose from day 0 to day 3, once daily. The drug dilutions of hydroxy functionalized compounds 7a-i and 10a-c were prepared in groundnut oil, while hemisuccinates 12a-i and 14a-c were dissolved in 5% NaHCO₃ to contain the required amount of the drug (0.6 mg for a dose of 48 mg/kg, 0.3 mg for a dose of 24 mg/kg, 0.15 mg for a dose of 12 mg/kg, 0.075 mg for a dose of 6 mg/kg, 0.00375 mg for a dose of 3 mg/kg) in 0.1 mL and administered intramuscular for each dose. Parasitemia levels were recorded from thin blood smears on day 4 and subsequently twice a week until day 28.11 The animals that did not develop patent infection until day 28 were recorded as cured. Multidrug-resistant Plasmodium yoelii nigeriensis used in this study is resistant to chloroquine, mefloquine, and halofantrine. Mice treated with β -arteether served as positive controls.

(10) (a) 100% suppression of parasitemia means no parasites were detected in 50 oil immersion microscopic fields (parasites, if at all present, are below the detection limit). The parasites present below the detection limit can multiply and eventually can be detected during observation on subsequent days. In such cases the drug is providing near 100% suppression of the parasitemia on day 4 but will not provide full protection to the treated mice in the 28 day survival assay. (b) 100% protection means none of the treated mice developed patent

infection during the 28 days of observation and hence were recorded as cured. Similarly 60% protection means only 3 out of 5 mice were cured.

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